



ISBN 978-1-960740-41-0

PHARMACOLOGY AND THERAPEUTICS – INNOVATIVE AND APPLIED CONCEPTS

Review Based Book Chapter

**PATHOPHYSIOLOGY AND TREATMENT APPROACHES FOR
AMYOTROPHIC LATERAL SCLEROSIS (ALS): A NARRATIVE
REVIEW**

October 09, 2024

doi: [10.5281/zenodo.13864703](https://doi.org/10.5281/zenodo.13864703)

Scientific Knowledge Publisher (SciKnowPub), USA
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REVIEW BASED BOOK CHAPTER**PATHOPHYSIOLOGY AND TREATMENT APPROACHES FOR AMYOTROPHIC LATERAL SCLEROSIS (ALS): A NARRATIVE REVIEW**J. Narayanan^{1*}, R. Sridevi¹, V. Chitra¹, V. Manimaran², T. Tamilanban¹¹Department of Pharmacology, SRM College of Pharmacy, SRM Institute of Science and Technology, Chennai, India²Department of Pharmaceutics, SRM College of Pharmacy, SRM Institute of Science and Technology, Chennai, India**For Correspondence**
narayanj@srmist.edu.in**Abstract**

A complicated neurodegenerative disease, amyotrophic lateral sclerosis (ALS) mostly affects motor neurons, resulting in a gradual weakening and atrophy of muscles. This review focuses into the pathophysiology of amyotrophic lateral sclerosis (ALS), providing insight on the complex genetic and molecular pathways at work and the ways in which it overlaps with frontotemporal dementia. Oxidative stress, excitotoxicity, mitochondrial dysfunction, disruption of axonal transport, neuroinflammation, DNA damage, and poor protein homeostasis are key pathological characteristic features of amyotrophic lateral sclerosis (ALS). Important hereditary factors include mutations in genes such as SOD1, FUS, and C9ORF72. About 15% of amyotrophic lateral sclerosis (ALS) patients also show signs of frontotemporal dementia, in addition to the characteristic motor symptoms and cognitive and behavioral abnormalities. The condition is characterized by aberrant TDP-43 or FUS protein aggregates, and the C9ORF72 gene mutation is the most prevalent genetic component. Physical therapy, dietary therapies, and respiratory support are all parts of the multidisciplinary care that is currently available to patients in an effort to reduce symptoms and enhance their quality of life. Gene therapies and novel pharmacological medicines that target specific disease pathways are among the novel treatments being researched. In order to address the complex nature of amyotrophic lateral sclerosis (ALS) and enhance patient outcomes, the field is shifting towards early intervention techniques and personalized medication.

Keywords

Amyotrophic Lateral Sclerosis (ALS), Frontotemporal Dementia, C9ORF72 Gene, Neuroinflammation, ALS Therapy

Introduction

A form of motor neuron disease that often manifests in adults, amyotrophic lateral sclerosis, is often called a neuromuscular complaint due to the fact that its symptoms are characterized by a reduction in muscle strength and size. The medical community has been under pressure due to advancements within the last two decades in the fields of clinical, genetic, and molecular biology. Similar to ALS with regard to cognitive, behavioural, and motor impairment, frontotemporal dementia is a unique kind of neurodegenerative illness. Fifteen percent of ALS patients exhibit frontotemporal dementia, while the remainder patients have varied levels of dysfunction in thinking or acting [1]. But in primary progressive aphasia (18%) and behavioural variant frontotemporal dementia (15%), ALS is present [2, 3]. In the cerebral cortex, inappropriate aggregates of transcription factor 43 (TDP-43) or fused in sarcoma (FUS) form in the cytoplasm of neurons. In individuals of European heritage, this can result in frontotemporal dementia, ALS, or both [4-6]. C9ORF72 is the most common gene involved in this process. Thus, amyotrophic lateral sclerosis (ALS) is commonly perceived as a complex neurodegenerative disease that falls within the spectrum of frontotemporal dementia-motor neuron disease (MND) [7, 8]. The recent development of amyotrophic lateral sclerosis (ALS) among this group of disorders has provided new possibilities for investigating the pathophysiology of the disease and developing novel therapeutic strategies.

Loss of both lower and higher motor neurons, alterations in muscle denervation, and degeneration of the corticobulbar/corticospinal pathways are the main symptoms of ALS. TDP-43 proteinopathy is involved in around 97% of cases of ALS, where TDP-43 forms cytoplasmic clumps after mislocalizing from the nucleus [9-11]. On the other hand, it is due to mutations in the cu-Zn superoxide dismutase 1 (SOD1) or FUS proteins exhibits distinct cytoplasmic protein aggregation and does not contain TDP-43 [12-15]. The most common genetic variant is characterized by excess p62-positive aggregates from dipeptide repeat proteins (DPRs) and TDP-43 mislocalization. These are linked to hexanucleotide expansions in C9orf72 [16].

A battery of blood tests, spine and brain imaging to eliminate out structural pathology, and a neurophysiological evaluation are among the diagnostic procedures that are ruled out due to mimic disorders of ALS [17]. For the diagnosis of ALS, the updated. The criteria of El Escorial [18], AWAJI Shima's specifications [19], in addition to the streamlined Gold Coast norms [20] were added. An ALS-prone patient is most affected by neuromuscular respiratory failure produced on by non-invasive ventilation [21, 22].

Enhancing the dosage used to cure ALS is the goal of cellular therapy [23]. By energy conservation and stretching exercises, physical therapy assists with problematic symptoms of ALS, such as fatigue and muscle stiffness [24]. It has been demonstrated that PT drastically improves an ALS patient's quality of life [25]. Various physical, psychological, and cognitive difficulties are faced by people with ALS, which can make it challenging to provide each aspect of their care. More research is needed to create nonpharmacological treatments for ALS that could lessen the emotional impact that the disease has on patients and those who care for them [26].

Epidemiology

Worldwide, ALS probability and occurrence were determined to be 4.42 and 1.59 per 100,000 people, respectively. Geographically, south Asia has the lowest and western Europe the highest. The incidence of ALS increases year by 0.00013, and more males than women are affected [27, 28].

PATHOPHYSIOLOGY

Genetic Makeup

In 1993, familial ALS (fALS) was estimated to be caused by SOD1 mutations in 10–21% of ALS patients. As of right now, 60–70% of cases are identified as fALS [29]. However, ALS is usually brought on by a combination of hereditary and environmental factors in 90–95% of cases. Genetic factors can cause ALS even in individuals without a family history, but the majority of cases (90–95%) are sporadic and have an estimated 50% heritability [30]. Genome-wide association studies have revealed that rare alterations have a significant role in shaping the molecular structure of ALS. The categorization of ALS has evolved from simple fALS and sALS to an enhanced complex risk gene-based molecular sub

classification [31]. Nearly thirty genes have been linked to ALS. In European families, a significant portion of cases, around 70%, are familial and have connections to FUS, C9ORF72, SOD1, and TARDBP. A recent study has uncovered 15 previously unknown risk loci [32], emphasizing the fact that ALS risk and disease severity are genetically independent. Neuroinflammation, mitochondrial dysfunction, and oxidative stress play crucial roles in physiological mechanisms. Every ALS patient has experienced positive outcomes from thorough genetic screening, as 21% of them have shown harmful mutations that can be clinically reported, while another 21% have variations of unclear significance [32, 33]. Genetic profiling has the potential to replace the outdated classifications of fALS and sALS, leading to advancements in therapy development and drug screening. The age of onset can vary even among members of the same family who have the same genetic mutation; in some cases, the mutation can exist for over 50 years prior to the beginning of disease. This indicates more than one process influenced due to a mixture of hereditary, behavioural, and ecological variables [34].

Oxidative Stress

Presently, significant existing research suggesting that vital significance of oxidative damage within the development of ALS, along with the compromised ability to defend against oxidative stress in the disease. This involves disrupting the cytoprotective system mediated by the antioxidant response element (ARE) on nuclear factor erythroid 2-related factor 2 (Nrf2), as well as glutathione homeostasis [35-40]. It has been demonstrated that both human ALS specimens and models of the illness exhibit changed oxidative stress biomarker profiles [41]. Aggregates of acetylated TDP-43 are enhanced by biochemical and cell-based methods, even though acetylation of TDP-43 prevents RNA binding and encourages the formation of hyperphosphorylated TDP-43 species, which are comparable to the harmful inclusions observed in the central nervous systems of ALS patients and in human ALS bio samples [42]. The phosphorylation of TDP-43 by GADD34 is enhanced by prolonged oxidative stress [43]. When TDP-43 aggregates in the cytoplasm, mitochondrial function is dysregulated [44]. Oxidative stress plays a role in the interaction between nearby astrocytes, microglia, and motor neurons [45, 46]. In a specific case, data suggests that astrocytes upregulate the

antiporter for cysteine and glutamate in reaction to elevated oxidative stress, which releases glutamate and can result in motor neuron excitotoxicity [47, 48].

Excitotoxicity

In ALS, excitotoxicity is caused by excessive postsynaptic glutamate receptor activation [49]. Prolonged elevation of glutamate at synapses in motor neuron excitotoxicity activation, increases amounts of calcium within cells, and inhibits the glutamate–cysteine antiporter, which lowers cysteine absorption. Long-term pathological alterations carried on by excitotoxicity include mitochondrial calcium excessive and ER stress [50-52]. In motor neurons that are affected by ALS, excitotoxic damage is more likely to occur because of decreased the expression of proteins that buffer calcium and increased expression of calcium-permeable AMPA receptors [53]. Metabolic glutamate receptors are becoming new potential therapeutic targets in ALS because of the possibility that their control will both reduce glutamate release and stimulate neurotrophic factor synthesis (NTFs).

Mitochondrial Dysfunction

Patient biosamples and ALS model systems describe disruptions to the mitochondria (axonal transport, structure, and dynamics), excessive production of ROS and decrease in ATP, disruption of calcium buffering, and induction of apoptosis [54, 55]. Several pathways have established a connection between mitochondrial dysfunction and mutations in certain amyotrophic lateral sclerosis (ALS) genes [56]. Numerous medications that target reactive oxygen species (ROS) and/or mitochondrial function, including creatine, coenzyme Q10, dexpramipexole, olesoxime, and dexpramipexole, performed well in animal models used for research but failed human trials [57].

Impaired Protein Homeostasis

Protein synthesis and degradation remain in control by a complex network that reacts to stress signals. This network includes the cytosolic heat shock response, unfolded protein responses in the endoplasmic reticulum (ER), and mitochondria [58]. Protein aggregates and a compromised proteome are hallmarks of age-related neurodegenerative disorders including spinal cord injury (the condition). Dysfunctional

proteostasis is a key component in the pathogenesis of amyotrophic lateral sclerosis (ALS), and several proteins associated with the disease either directly or indirectly control proteostasis [58-60]. Specifically, maturation is regulated by ubiquitin 2, alsin, FIG4, VCP, and CHMP2B, whereas autophagy initiation is dependent on ubiquitin 2, optineurin, sequestosome 1/P62, and C9ORF72 [61]. Motor neurons are particularly vulnerable to proteome stress due to their large size, minimal heat shock response, and restricted expression of ubiquitin proteasome genes. One of the main characteristics of ALS is intracellular protein aggregates, particularly TDP-43 mislocalization, which causes abnormal splicing, increases DNA damage, changes the transcriptome, and impairs axonal translation [62, 63]. UNC13A mRNA has cryptic exon inclusion due to TDP-43 loss from the nucleus, which lowers UNC13A protein expression and may have an impact on vesicle development and neurotransmitter release [64, 65].

Neuroinflammation and Glial Contribution

Inflammation of the nervous system or brain pathogen-induced cause of the preclinical stage of ALS [66]. Astrocytes control inflammatory signalling and maintain the integrity of the blood-brain barrier by the production of either pro- or anti-inflammatory cytokines, such as prostaglandin E2 and transforming growth factor (TGF)- β [67-69]. Motor neurons cocultured with astrocytes generated from fibroblasts from ALS patients are hazardous to them [70]. The specific ways this toxicity occurs are still unknown, however decreased bioenergetic support due to lactate release and activation of the pro-nerve growth factor-p75 receptor are possible causes [71]. One major process of neuroinflammation in amyotrophic lateral sclerosis (ALS) is the activation of the microglial NLRP3 inflammasome. Regarding its possible use as a therapeutic target, NLRP3 inhibition has one that potential to decrease microglia-induced neuroinflammation and halt the progression of amyotrophic lateral sclerosis (ALS). The NF- κ B protein is a master regulator of inflammation in persons with amyotrophic lateral sclerosis (ALS), according to recent discoveries [72].

DNA Damage and Repair

Oxidized deoxyguanosine (OdG) levels are greater in neurons and other postmitotic cells in central nervous system (CNS) tissues and biofluids from amyotrophic lateral

sclerosis (ALS) patients [73, 74]. New research has connected amyotrophic lateral sclerosis (ALS) to DNA damage response (DDR) activation and an increase in apurinic/aprimidinic DNA sites, which are areas where DNA bases are damaged [75].

Impaired Axonal Transport and Integrity

In many models of amyotrophic lateral sclerosis (ALS) and across human patients, pathological buildups of organelles and phosphorylated neurofilaments within the terminals of motor neurons have been associated to defects in axonal transport [76]. New evidence suggests that amyotrophic lateral sclerosis (ALS) is the result of genetic mutations ANXA11 (a protein that hinders axonal RNA transport) and KIF5A (a protein that encodes a microtubule motor) [77, 78]. In SOD1-mutant rats, P38 MAPK inhibitors reverted back normal axonal transport, but in a targeted manner, IGF1R inhibitors improved the axonal transport of signalling endosomes. The presence of signalling endosomes in retrograde axonal transit is essential for maintaining axonal integrity [79, 80].

Advances in ALS Therapy

Multidisciplinary Care

American amyotrophic lateral sclerosis (ALS) patients may see a multidisciplinary team of healthcare professionals during a single clinic visit, including a pulmonologist, a speech-a social worker, a dietitian, a linguist, a physiotherapist, an occupational therapist, and an ALS specialist. An integrative strategy is used in the treatment of ALS. Connections to ALS/MND groups are made in order to obtain further assistance [81, 82].

Modulators of Disease

Excitotoxicity, oxidative stress, mitochondrial dysfunction, protein homeostasis, neuroinflammation, cell death, cytoskeletal integrity, axonal transport, DNA repair, RNA metabolism, and stress granule modulation have been the primary areas of emphasis for ALS therapy in the last 20 years [83, 84]. To ensure the effective development of ALS therapy, the worldwide ALS community must prioritize future initiatives such as improving data exchange, endpoint harmonization, and trial design and analysis and ensuring

equity of access. A treatment for amyotrophic lateral sclerosis (ALS) has been approved by the US Food and Drug Administration (FDA): a combination of sodium phenylbutyrate and taurursodiol. This medication targets mitochondrial malfunction, endoplasmic reticulum stress, and cell death [85]. Other approved medications include riluzole, an anti-glutamine medication [86], and edaravone, which decreases oxidative stress [87] does not have European approval with the purpose of addressing amyotrophic lateral sclerosis [88, 89].

Pulmonary Intervention

As mentioned earlier, the main cause of death for ALS patients is respiratory failure. For the treatment of amyotrophic lateral sclerosis (ALS), it is essential to do pulmonary tests such as spirometry, polysomnography, transdiaphragmatic pressure, sniff nasal pressure, arterial blood gas, and nocturnal pulse oximetry. Regular evaluations are crucial for spotting respiratory muscle weakness and facilitating early non-invasive ventilation intervention, both of which can improve survival and quality of life. Even though there isn't much thorough research on the advantages of mechanical insufflation-exsufflation, it's commonly used to help people cough and clear their airways. Research on integrating respiratory muscle training with swallowing exercises to improve coughing and swallowing is still ongoing [90, 91].

Diet and Nutritional Intervention

Reduction in ALSFRS-R scores is associated with a decrease in body mass [92]. Losing weight is complicated and associated with several problems, such as fatigue, hyper metabolism, dysphagia, reduced meal intake, and impaired dexterity of the limbs when using utensils [21]. A shorter survival time was linked to body mass index extremes (<18, >40), while the 30-35 body mass index range showed the best survival [93]. Most agrees that antioxidants, fruits, fiber, and carotenes should be included in the diet [94]. According to clinical guidelines, patients who have symptomatic dysphagia, prolonged feeding times, severe weight loss (more than 5–10%), and, in some situations, deteriorating respiratory function, should be evaluated for gastrostomy tube placement. Proper treatment, procedure scheduling, and patient selection are critical to the success of a gastrostomy. An excellent resource for ALS patients, caregivers, and

doctors, the Simplifying ALS initiative (www.alsuntangled.com) has evaluated the data for numerous dietary supplements and vitamins. Nevertheless, the majority of clinical trials have not shown a lower incidence of ALS progression [95, 96]. Studies have shown that vitamin E may protect against amyotrophic lateral sclerosis (ALS), and a recent phase 3 experiment found that treating patients with an ultrahigh dose of methylcobalamin (50 mg) somewhat slowed their clinical deterioration compared to placebo [56, 97, 98].

Emerging Treatments

An increase in ALS research treatments has occurred due to the ineffectiveness of FDA-approved drugs. The ALS platform trial makes use of a centralized infrastructure and a shared master protocol, enabling the simultaneous evaluation of several agents [99]. At least fifty tiny compounds are being studied for various uses. The FDA-approved medication tofersen (2023) has demonstrated efficacy in treating pathogenic gene expression in SOD1-ALS patients. Phase 1 studies are being conducted on gene therapy vectors that use adeno-associated viruses to reduce SOD1 levels, while phase 1-2 trials are now being conducted on antisense oligonucleotides that target C9ORF72 and FUS. Phase 2 trials are also evaluating monoclonal antibodies that target misfolded proteins.

Conclusion

Over the past twenty years, we have seen a significant advancement in basic ALS research. But one of the main reasons for patient and caregiver frustration is the absence of substantial progress in converting this amount of knowledge into practical treatments. By consolidating biomarkers, developing systematic and innovative clinical trials, and concentrating on early disease among carriers of pre-symptomatic gene mutations, the discipline is drawing nearer to translating fundamental scientific discoveries into treatments that affect disease. As with cancer therapeutics, the hope is that the larger population of people living with sporadic amyotrophic lateral sclerosis (ALS) can find a personalized solution that limits disability and allows them to live with dignity by identifying risk gene variants and finding ways to identify the dominant mechanism (e.g., pathogenic inflammation, activation of retro-transposons, and oxidative stress) serving as the agent responsible for spreading some diseases.

Conflicts of Interest

The authors declared no conflict of interest.

References

- [1] Phukan, J., Elamin, M., Bede, P., Jordan, N., Gallagher, L., Byrne, S., ... & Hardiman, O. (2012). The syndrome of cognitive impairment in amyotrophic lateral sclerosis: a population-based study. *Journal of Neurology, Neurosurgery & Psychiatry*, 83(1), 102-108.
- [2] Lomen-Hoerth, C., Anderson, T., & Miller, B. (2002). The overlap of amyotrophic lateral sclerosis and frontotemporal dementia. *Neurology*, 59(7), 1077-1079.
- [3] Rascovsky, K., Hodges, J. R., Knopman, D., Mendez, M. F., Kramer, J. H., Neuhaus, J., ... & Miller, B. L. (2011). Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia. *Brain*, 134(9), 2456-2477.
- [4] Renton, A. E., Majounie, E., Waite, A., Simón-Sánchez, J., Rollinson, S., Gibbs, J. R., ... & Traynor, B. J. (2011). A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron*, 72(2), 257-268.
- [5] Neumann, M., Sampathu, D. M., Kwong, L. K., Truax, A. C., Micsenyi, M. C., Chou, T. T., ... & Lee, V. M. Y. (2006). Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*, 314(5796), 130-133.
- [6] Deng, H. X., Zhai, H., Bigio, E. H., Yan, J., Fecto, F., Ajroud, K., ... & Siddique, T. (2010). FUS-immunoreactive inclusions are a common feature in sporadic and non-SOD1 familial amyotrophic lateral sclerosis. *Annals of Neurology*, 67(6), 739-748.
- [7] Hardiman, O., Al-Chalabi, A., Chio, A., Corr, E. M., Logroscino, G., Robberecht, W., ... & Van Den Berg, L. H. (2017). Amyotrophic lateral sclerosis. *Nature Reviews Disease Primers*, 3(1), 1-19.
- [8] Burrell, J. R., Halliday, G. M., Kril, J. J., Ittner, L. M., Götz, J., Kiernan, M. C., & Hodges, J. R. (2016). The frontotemporal dementia-motor neuron disease continuum. *The Lancet*, 388(10047), 919-931.
- [9] Scotter, E. L., Chen, H. J., & Shaw, C. E. (2015). TDP-43 proteinopathy and ALS: insights into disease mechanisms and therapeutic targets. *Neurotherapeutics*, 12(2), 352-363.
- [10] Ling, S. C., Polymenidou, M., & Cleveland, D. W. (2013). Converging mechanisms in ALS and FTD: disrupted RNA and protein homeostasis. *Neuron*, 79(3), 416-438.
- [11] Tan, R. H., Ke, Y. D., Ittner, L. M., & Halliday, G. M. (2017). ALS/FTLD: experimental models and reality. *Acta Neuropathologica*, 133(2), 177-196.
- [12] Gu, S., Xu, M., Chen, L., Shi, X., & Luo, S. Z. (2023). A liquid-to-solid phase transition of Cu/Zn superoxide dismutase 1 initiated by oxidation and disease mutation. *Journal of Biological Chemistry*, 299(2).
- [13] Mackenzie, I. R., Bigio, E. H., Ince, P. G., Geser, F., Neumann, M., Cairns, N. J., ... & Trojanowski, J. Q. (2007). Pathological TDP-43 distinguishes sporadic amyotrophic lateral sclerosis from amyotrophic lateral sclerosis with SOD1 mutations. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*, 61(5), 427-434.
- [14] Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., ... & Shaw, C. E. (2009). Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*, 323(5918), 1208-1211.
- [15] Kato, S., Sumi-Akamaru, H., Fujimura, H., Sakoda, S., Kato, M., Hirano, A., ... & Ohama, E. (2001). Copper chaperone for superoxide dismutase co-aggregates with superoxide dismutase 1 (SOD1) in neuronal Lewy body-like hyaline inclusions: an immunohistochemical study on familial amyotrophic lateral sclerosis with SOD1 gene mutation. *Acta Neuropathologica*, 102, 233-238.
- [16] Ramos-Campoy, O., Ávila-Polo, R., Grau-Rivera, O., Antonell, A., Clarimón, J., Rojas-García, R., ... & Gelpi, E. (2018). Systematic screening of ubiquitin/p62 aggregates in cerebellar cortex expands the neuropathological phenotype of the C9orf72 expansion mutation. *Journal of Neuropathology & Experimental Neurology*, 77(8), 703-709.

- [17] Turner, M. R., & Talbot, K. (2013). Mimics and chameleons in motor neurone disease. *Practical Neurology*, 13(3), 153-164.
- [18] Brooks, B. R., Miller, R. G., Swash, M., & Munsat, T. L. (2000). El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 1(5), 293-299.
- [19] De Carvalho, M., Dengler, R., Eisen, A., England, J. D., Kaji, R., Kimura, J., ... & Swash, M. (2008). Electrodiagnostic criteria for diagnosis of ALS. *Clinical Neurophysiology*, 119(3), 497-503.
- [20] Shefner, J. M., Al-Chalabi, A., Baker, M. R., Cui, L. Y., de Carvalho, M., Eisen, A., ... & Kiernan, M. C. (2020). A proposal for new diagnostic criteria for ALS. *Clinical Neurophysiology*, 131(8), 1975-1978.
- [21] Bourke, S. C., Tomlinson, M., Williams, T. L., Bullock, R. E., Shaw, P. J., & Gibson, G. J. (2006). Effects of non-invasive ventilation on survival and quality of life in patients with amyotrophic lateral sclerosis: a randomised controlled trial. *The Lancet Neurology*, 5(2), 140-147.
- [22] Lechtzin, N., Scott, Y., Busse, A. M., Clawson, L. L., Kimball, R., & Wiener, C. M. (2007). Early use of non-invasive ventilation prolongs survival in subjects with ALS. *Amyotrophic Lateral Sclerosis*, 8(3), 185-188.
- [23] Lin, T. J., Cheng, K. C., Wu, L. Y., Lai, W. Y., Ling, T. Y., Kuo, Y. C., & Huang, Y. H. (2022). Potential of cellular therapy for ALS: current strategies and future prospects. *Frontiers in Cell and Developmental Biology*, 10, 851613.
- [24] Masrori, P., & Van Damme, P. (2020). Amyotrophic lateral sclerosis: a clinical review. *European Journal of Neurology*, 27(10), 1918-1929.
- [25] Tzeplaeff, L., Wilfling, S., Requardt, M. V., & Herdick, M. (2023). Current state and future directions in the therapy of ALS. *Cells*, 12(11), 1523.
- [26] Meyer, R., Spittel, S., Steinfurth, L., Funke, A., Kettemann, D., Münch, C., ... & Maier, A. (2018). Patient-reported outcome of physical therapy in amyotrophic lateral sclerosis: observational online study. *JMIR Rehabilitation and Assistive Technologies*, 5(2), e10099.
- [27] Arthur, K. C., Calvo, A., Price, T. R., Geiger, J. T., Chio, A., & Traynor, B. J. (2016). Projected increase in amyotrophic lateral sclerosis from 2015 to 2040. *Nature Communications*, 7(1), 12408.
- [28] Xu, L., Liu, T., Liu, L., Yao, X., Chen, L., Fan, D., ... & Wang, S. (2020). Global variation in prevalence and incidence of amyotrophic lateral sclerosis: a systematic review and meta-analysis. *Journal of Neurology*, 267, 944-953.
- [29] Ranganathan, R., Haque, S., Coley, K., Shephard, S., Cooper-Knock, J., & Kirby, J. (2020). Multifaceted genes in amyotrophic lateral sclerosis-frontotemporal dementia. *Frontiers in Neuroscience*, 14, 684.
- [30] Van Rheenen, W., Van Der Spek, R. A., Bakker, M. K., Van Vugt, J. J., Hop, P. J., Zwamborn, R. A., ... & Mathers, S. (2021). Common and rare variant association analyses in amyotrophic lateral sclerosis identify 15 risk loci with distinct genetic architectures and neuron-specific biology. *Nature Genetics*, 53(12), 1636-1648.
- [31] Van Rheenen, W., Shatunov, A., Dekker, A. M., McLaughlin, R. L., Diekstra, F. P., Pulit, S. L., ... & Kurth, I. (2016). Genome-wide association analyses identify new risk variants and the genetic architecture of amyotrophic lateral sclerosis. *Nature Genetics*, 48(9), 1043-1048.
- [32] Shephard, S. R., Parker, M. D., Cooper-Knock, J., Verber, N. S., Tuddenham, L., Heath, P., ... & Shaw, P. J. (2021). Value of systematic genetic screening of patients with amyotrophic lateral sclerosis. *Journal of Neurology, Neurosurgery & Psychiatry*, 92(5), 510-518.
- [33] Zhang, S., Cooper-Knock, J., Weimer, A. K., Shi, M., Moll, T., Marshall, J. N., ... & Snyder, M. P. (2022). Genome-wide identification of the genetic basis of amyotrophic lateral sclerosis. *Neuron*, 110(6), 992-1008.
- [34] Al-Chalabi, A., Calvo, A., Chio, A., Colville, S., Ellis, C. M., Hardiman, O., ... & Pearce, N. (2014). Analysis of amyotrophic lateral sclerosis as a multistep process: a population-based modelling study. *The Lancet Neurology*, 13(11), 1108-1113.

- [35] Mitsumoto, H., Santella, R. M., Liu, X., Bogdanov, M., Zipprich, J., Wu, H. C., ... & Factor-Litvak, P. (2008). Oxidative stress biomarkers in sporadic ALS. *Amyotrophic Lateral Sclerosis*, 9(3), 177-183.
- [36] Kim, K. (2021). Glutathione in the nervous system as a potential therapeutic target to control the development and progression of amyotrophic lateral sclerosis. *Antioxidants*, 10(7), 1011.
- [37] Cuadrado, A., Rojo, A. I., Wells, G., Hayes, J. D., Cousin, S. P., Rumsey, W. L., ... & Dinkova-Kostova, A. T. (2019). Therapeutic targeting of the NRF2 and KEAP1 partnership in chronic diseases. *Nature Reviews Drug Discovery*, 18(4), 295-317.
- [38] Jiménez-Villegas, J., Ferraiuolo, L., Mead, R. J., Shaw, P. J., Cuadrado, A., & Rojo, A. I. (2021). NRF2 as a therapeutic opportunity to impact in the molecular roadmap of ALS. *Free Radical Biology and Medicine*, 173, 125-141.
- [39] Cohen, T. J., Hwang, A. W., Restrepo, C. R., Yuan, C. X., Trojanowski, J. Q., & Lee, V. M. (2015). An acetylation switch controls TDP-43 function and aggregation propensity. *Nature Communications*, 6(1), 5845.
- [40] Goh, C. W., Lee, I. C., Sundaram, J. R., George, S. E., Yusoff, P., Brush, M. H., ... & Shenolikar, S. (2018). Chronic oxidative stress promotes GADD34-mediated phosphorylation of the TAR DNA-binding protein TDP-43, a modification linked to neurodegeneration. *Journal of Biological Chemistry*, 293(1), 163-176.
- [41] Zuo, X., Zhou, J., Li, Y., Wu, K., Chen, Z., Luo, Z., ... & Fu, X. D. (2021). TDP-43 aggregation induced by oxidative stress causes global mitochondrial imbalance in ALS. *Nature Structural & Molecular Biology*, 28(2), 132-142.
- [42] Kazama, M., Kato, Y., Kakita, A., Noguchi, N., Urano, Y., Masui, K., ... & Shibata, N. (2020). Astrocytes release glutamate via cystine/glutamate antiporter upregulated in response to increased oxidative stress related to sporadic amyotrophic lateral sclerosis. *Neuropathology*, 40(6), 587-598.
- [43] Deora, V., Lee, J. D., Albornoz, E. A., McAlary, L., Jagaraj, C. J., Robertson, A. A., ... & Woodruff, T. M. (2020). The microglial NLRP3 inflammasome is activated by amyotrophic lateral sclerosis proteins. *Glia*, 68(2), 407-421.
- [44] King, A. E., Woodhouse, A., Kirkcaldie, M. T., & Vickers, J. C. (2016). Excitotoxicity in ALS: overstimulation, or overreaction?. *Experimental Neurology*, 275, 162-171.
- [45] Van Cutsem, P., Dewil, M., Robberecht, W., & Van Den Bosch, L. (2006). Excitotoxicity and amyotrophic lateral sclerosis. *Neurodegenerative Diseases*, 2(3-4), 147-159.
- [46] Lewerenz, J., Hewett, S. J., Huang, Y., Lambros, M., Gout, P. W., Kalivas, P. W., ... & Maher, P. (2013). The cystine/glutamate antiporter system xc⁻ in health and disease: from molecular mechanisms to novel therapeutic opportunities. *Antioxidants and Redox Signaling*, 18(5), 522-555.
- [47] Williams, T. L., Day, N. C., Ince, P. G., Kamboj, R. K., & Shaw, P. J. (1997). Calcium-permeable α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptors: A molecular determinant of selective vulnerability in amyotrophic lateral sclerosis. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*, 42(2), 200-207.
- [48] Ince, P., Stout, N., Shaw, P., Slade, J., Hunziker, W., Heizmann, C. W., & Baimbridge, K. G. (1993). Parvalbumin and calbindin D-28k in the human motor system and in motor neuron disease. *Neuropathology and Applied Neurobiology*, 19(4), 291-299.
- [49] Cabral-Costa, J. V., & Kowaltowski, A. J. (2020). Neurological disorders and mitochondria. *Molecular Aspects of Medicine*, 71, 100826.
- [50] Jhanji, R., Behl, T., Sehgal, A., & Bungau, S. (2021). Mitochondrial dysfunction and traffic jams in amyotrophic lateral sclerosis. *Mitochondrion*, 58, 102-110.
- [51] Debska-Vielhaber, G., Miller, I., Peeva, V., Zuschratter, W., Walczak, J., Schreiber, S., ... & Kunz, W. S. (2021). Impairment of mitochondrial oxidative phosphorylation in skin fibroblasts of SALS and FALS patients is rescued by in vitro treatment with ROS scavengers. *Experimental Neurology*, 339, 113620.

- [52] Bernard-Marissal, N., Chrast, R., & Schneider, B. L. (2018). Endoplasmic reticulum and mitochondria in diseases of motor and sensory neurons: a broken relationship?. *Cell Death & Disease*, 9(3), 333.
- [54] Cudkowicz, M. E., van den Berg, L. H., Shefner, J. M., Mitsumoto, H., Mora, J. S., Ludolph, A., ... & Kerr, D. A. (2013). Dexamipexole versus placebo for patients with amyotrophic lateral sclerosis (EMPOWER): a randomised, double-blind, phase 3 trial. *The Lancet Neurology*, 12(11), 1059-1067.
- [55] Pastula, D. M., Moore, D. H., & Bedlack, R. S. (2012). Creatine for amyotrophic lateral sclerosis/motor neuron disease. *Cochrane Database of Systematic Reviews*, (12).
- [56] Kaufmann, P., Thompson, J. L., Levy, G., Buchsbaum, R., Shefner, J., Krivickas, L. S., ... & Levin, B. (2009). Phase II trial of CoQ10 for ALS finds insufficient evidence to justify phase III. *Annals of Neurology: Official Journal of the American Neurological Association and the Child Neurology Society*, 66(2), 235-244.
- [57] Lenglet, T., Lacomblez, L., Abitbol, J. L., Ludolph, A., Mora, J. S., Robberecht, W., ... & Hanisch, F. (2014). A phase II– III trial of olesoxime in subjects with amyotrophic lateral sclerosis. *European Journal of Neurology*, 21(3), 529-536.
- [58] Webster, C. P., Smith, E. F., Shaw, P. J., & De Vos, K. J. (2017). Protein homeostasis in amyotrophic lateral sclerosis: therapeutic opportunities?. *Frontiers in Molecular Neuroscience*, 10, 123.
- [59] Ramesh, N., & Pandey, U. B. (2017). Autophagy dysregulation in ALS: when protein aggregates get out of hand. *Frontiers in Molecular Neuroscience*, 10, 263.
- [60] Montibeller, L., Tan, L. Y., Kim, J. K., Paul, P., & de Belleruche, J. (2020). Tissue-selective regulation of protein homeostasis and unfolded protein response signalling in sporadic ALS. *Journal of Cellular and Molecular Medicine*, 24(11), 6055-6069.
- [61] Yerbury, J. J., Farrawell, N. E., & McAlary, L. (2020). Proteome homeostasis dysfunction: a unifying principle in ALS pathogenesis. *Trends in Neurosciences*, 43(5), 274-284.
- [62] Suk, T. R., & Rousseaux, M. W. (2020). The role of TDP-43 mislocalization in amyotrophic lateral sclerosis. *Molecular Neurodegeneration*, 15(1), 45.
- [63] Mitra, J., Guerrero, E. N., Hegde, P. M., Liachko, N. F., Wang, H., Vasquez, V., ... & Hegde, M. L. (2019). Motor neuron disease-associated loss of nuclear TDP-43 is linked to DNA double-strand break repair defects. *Proceedings of the National Academy of Sciences*, 116(10), 4696-4705.
- [64] Nagano, S., Jinno, J., Abdelhamid, R. F., Jin, Y., Shibata, M., Watanabe, S., ... & Araki, T. (2020). TDP-43 transports ribosomal protein mRNA to regulate axonal local translation in neuronal axons. *Acta Neuropathologica*, 140, 695-713.
- [65] Brown, A. L., Wilkins, O. G., Keuss, M. J., Hill, S. E., Zanovello, M., Lee, W. C., ... & Fratta, P. (2022). TDP-43 loss and ALS-risk SNPs drive mis-splicing and depletion of UNC13A. *Nature*, 603(7899), 131-137.
- [66] Appel, S. H., Beers, D. R., & Zhao, W. (2021). Amyotrophic lateral sclerosis is a systemic disease: peripheral contributions to inflammation-mediated neurodegeneration. *Current Opinion in Neurology*, 34(5), 765-772.
- [67] Vu, L., An, J., Kovalik, T., Gendron, T., Petrucelli, L., & Bowser, R. (2020). Cross-sectional and longitudinal measures of chitinase proteins in amyotrophic lateral sclerosis and expression of CHI3L1 in activated astrocytes. *Journal of Neurology, Neurosurgery & Psychiatry*, 91(4), 350-358.
- [68] Westergard, T., & Rothstein, J. D. (2020). Astrocyte diversity: current insights and future directions. *Neurochemical Research*, 45(6), 1298-1305.
- [69] Yamanaka, K., & Komine, O. (2018). The multi-dimensional roles of astrocytes in ALS. *Neuroscience Research*, 126, 31-38.
- [70] Meyer, K., Ferraiuolo, L., Miranda, C. J., Likhite, S., McElroy, S., Renssch, S., ... & Kaspar, B. K. (2014). Direct conversion of patient fibroblasts demonstrates non-cell autonomous toxicity of astrocytes to motor neurons in familial and sporadic ALS. *Proceedings of the National Academy of Sciences*, 111(2), 829-832.

- [71] Haidet-Phillips, A. M., Hester, M. E., Miranda, C. J., Meyer, K., Braun, L., Frakes, A., ... & Kaspar, B. K. (2011). Astrocytes from familial and sporadic ALS patients are toxic to motor neurons. *Nature Biotechnology*, 29(9), 824-828.
- [72] Yu, C. H., Davidson, S., Harapas, C. R., Hilton, J. B., Mlodzianoski, M. J., Laohamonthonkul, P., ... & Masters, S. L. (2020). TDP-43 triggers mitochondrial DNA release via mPTP to activate cGAS/STING in ALS. *Cell*, 183(3), 636-649.
- [73] Kok, J. R., Palminha, N. M., Dos Santos Souza, C., El-Khamisy, S. F., & Ferraiuolo, L. (2021). DNA damage as a mechanism of neurodegeneration in ALS and a contributor to astrocyte toxicity. *Cellular and Molecular Life Sciences*, 78(15), 5707-5729.
- [74] Ferrante, R. J., Browne, S. E., Shinobu, L. A., Bowling, A. C., Baik, M. J., MacGarvey, U., ... & Beal, M. F. (1997). Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *Journal of Neurochemistry*, 69(5), 2064-2074.
- [75] Bogdanov, M., Brown Jr, R. H., Matson, W., Smart, R., Hayden, D., O'Donnell, H., ... & Cudkovicz, M. (2000). Increased oxidative damage to DNA in ALS patients. *Free Radical Biology and Medicine*, 29(7), 652-658.
- [76] De Vos, K. J., Grierson, A. J., Ackerley, S., & Miller, C. C. (2008). Role of axonal transport in neurodegenerative diseases. *Annual Review of Neuroscience*, 31(1), 151-173.
- [77] Nicolas, A., Kenna, K. P., Renton, A. E., Ticozzi, N., Faghri, F., Chia, R., ... & Cooper, G. M. (2018). Genome-wide analyses identify KIF5A as a novel ALS gene. *Neuron*, 97(6), 1267-1288.
- [78] Liao, Y. C., Fernandopulle, M. S., Wang, G., Choi, H., Hao, L., Drerup, C. M., ... & Ward, M. E. (2019). RNA granules hitchhike on lysosomes for long-distance transport, using annexin A11 as a molecular tether. *Cell*, 179(1), 147-164.
- [79] Gibbs, K. L., Kalmar, B., Rhymes, E. R., Fellows, A. D., Ahmed, M., Whiting, P., ... & Schiavo, G. (2018). Inhibiting p38 MAPK alpha rescues axonal retrograde transport defects in a mouse model of ALS. *Cell Death & Disease*, 9(6), 596.
- [80] Fellows, A. D., Rhymes, E. R., Gibbs, K. L., Greensmith, L., & Schiavo, G. (2020). IGF 1R regulates retrograde axonal transport of signalling endosomes in motor neurons. *EMBO Reports*, 21(3), e49129.
- [81] Miller RG, Jackson CE, Kasarskis EJ, et al, Quality Standards Subcommittee of the American Academy of Neurology. Practice parameter update: the care of the patient with amyotrophic lateral sclerosis: multidisciplinary care, symptom management, and cognitive/behavioral impairment (an evidence-based review): report of the Quality Standards Subcommittee of the American Academy of Neurology. (2009). *Neurology*, 73:1227-33.
- [82] EFNS Task Force on Diagnosis and Management of Amyotrophic Lateral Sclerosis:, Andersen, P. M., Abrahams, S., Borasio, G. D., de Carvalho, M., Chio, A., ... & Weber, M. (2012). EFNS guidelines on the clinical management of amyotrophic lateral sclerosis (MALS)—revised report of an EFNS task force. *European Journal of Neurology*, 19(3), 360-375.
- [83] Chiò, A., Mazzini, L., & Mora, G. (2020). Disease-modifying therapies in amyotrophic lateral sclerosis. *Neuropharmacology*, 167, 107986.
- [84] Bensimon, G., Lacomblez, L., Meininger, V. A. L. S., & ALS/Riluzole Study Group. (1994). A controlled trial of riluzole in amyotrophic lateral sclerosis. *New England Journal of Medicine*, 330(9), 585-591.
- [85] Glass, J. D., & Fournier, C. N. (2022). Unintended consequences of approving unproven treatments—hope, hype, or harm?. *JAMA Neurology*, 79(2), 117-118.
- [86] Group II, R. S., Lacomblez, L., Bensimon, G., Meininger, V., Leigh, P. N., & Guillet, P. (1996). Dose-ranging study of riluzole in amyotrophic lateral sclerosis. *The Lancet*, 347(9013), 1425-1431.
- [87] Writing Group, Edaravone (MCI-186) ALS 19 Study Group. Safety and efficacy of edaravone in well defined patients with amyotrophic lateral sclerosis: a randomised, double-blind, placebo-controlled trial. (2017). *Lancet Neurology*, 2017,16:505-12.
- [88] Takahashi, F., Takei, K., Tsuda, K., & Palumbo, J. (2017). Post-hoc analysis of MCI186-17, the extension study to MCI186-16, the confirmatory double-blind, parallel-group, placebo-controlled

study of edaravone in amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration*, 18(sup1), 32-39.

[89] Al-Chalabi, A., Andersen, P. M., Chandran, S., Chio, A., Corcia, P., Couratier, P., ... & Van Den Berg, L. H. (2017). July 2017 ENCALS statement on edaravone. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration*, 18(7-8), 471-474.

[90] Paganoni, S., Macklin, E. A., Hendrix, S., Berry, J. D., Elliott, M. A., Maiser, S., ... & Cudkovicz, M. E. (2020). Trial of sodium phenylbutyrate–taurursodiol for amyotrophic lateral sclerosis. *New England Journal of Medicine*, 383(10), 919-930.

[91] Miller, R. G., Jackson, C. E., Kasarskis, E. J., England, J. D., Forshew, D., Johnston, W., ... & Woolley, S. C. (2009). Practice Parameter update: the care of the patient with amyotrophic lateral sclerosis: drug, nutritional, and respiratory therapies (an evidence-based review) Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology*, 73(15), 1218-1226.

[92] Lechtzin, N., Cudkovicz, M. E., de Carvalho, M., Genge, A., Hardiman, O., Mitsumoto, H., ... & Andrews, J. A. (2018). Respiratory measures in amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration*, 19(5-6), 321-330.

[93] Plowman, E. K., Watts, S. A., Tabor, L., Robison, R., Gaziano, J., Domer, A. S., ... & Gooch, C. (2016). Impact of expiratory strength training in amyotrophic lateral sclerosis. *Muscle & Nerve*, 54(1), 48-53.

[94] Lee, I., Kazamel, M., McPherson, T., McAdam, J., Bamman, M., Amara, A., ... & King, P. H. (2021). Fat mass loss correlates with faster disease progression in amyotrophic lateral sclerosis patients: Exploring the utility of dual-energy x-ray absorptiometry in a prospective study. *PLoS One*, 16(5), e0251087.

[95] Dupuis, L., Pradat, P. F., Ludolph, A. C., & Loeffler, J. P. (2011). Energy metabolism in amyotrophic lateral sclerosis. *The Lancet Neurology*, 10(1), 75-82.

[96] Paganoni, S., Deng, J., Jaffa, M., Cudkovicz, M. E., & Wills, A. M. (2011). Body mass index, not dyslipidemia, is an independent predictor of survival in amyotrophic lateral sclerosis. *Muscle & Nerve*, 44(1), 20-24.

[97] Prell, T., Grosskreutz, J., & Pooled Resource Open-Access ALS Clinical Trials Consortium. (2020). Use of vitamins by participants in amyotrophic lateral sclerosis clinical trials. *Plos One*, 15(8), e0237175.

[98] Wang, H., O'Reilly, É. J., Weisskopf, M. G., Logroscino, G., McCullough, M. L., Schatzkin, A., ... & Ascherio, A. (2011). Vitamin E intake and risk of amyotrophic lateral sclerosis: a pooled analysis of data from 5 prospective cohort studies. *American Journal of Epidemiology*, 173(6), 595-602.

[99] Oki, R., Izumi, Y., Fujita, K., Miyamoto, R., Nodera, H., Sato, Y., ... & Fujino, Y. (2022). Efficacy and safety of ultrahigh-dose methylcobalamin in early-stage amyotrophic lateral sclerosis: a randomized clinical trial. *JAMA Neurology*, 79(6), 575-583.

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